

No.: PMEB-1/Spl./10(2)/2023-24

Date: 06-01-2024

NOTIFICATION

Sub.: Syllabus and Examination pattern of **M.Sc. (Reproductive Biology & Clinical Embryology-IVF)** course under Specialized Programmes from the academic year 2023-24-reg.

- Ref.: 1. Decision of the BOS Meeting held on 03-06-2023.
2. Decision of the Academic Council meeting held on 10-11-2023.

The Board of Studies in **M.Sc. (Reproductive Biology & Clinical Embryology-IVF) (PG)** at its meeting held on 03-06-2023 has recommended to approve the scheme of examination and the syllabus of **M.Sc. (Reproductive Biology & Clinical Embryology-IVF)** course in University of Mysore under specialized/specified programs as per CBCS Scheme from the academic year 2023-24.

The Academic Council has also approved the above said proposals at its meeting held on 10-11-2023 and the same is hereby notified.

The syllabus of **M.Sc. (Reproductive Biology & Clinical Embryology-IVF)** course may be downloaded from the University website <https://uni-mysore.ac.in/PMEB/>.


REGISTRAR

REGISTRAR

University of Mysore

MYSURU - 570 005

To,

1. The Registrar (Evaluation), University of Mysore, Mysuru.
2. The Dean, Faculty of Science & Technology, DoS in Earth Science, Manasagangothri, Mysuru.
3. Prof. Kemparaju K, DoS in Bio chemistry, Manasagangothri, Mysuru.
4. The Principal, Cauvery College of Life Science & Management, #131/1, 2D, 2E, 2F, KBL Layout, Near Devegowda Circle, Alanahalli, Mysuru.
5. The Deputy Registrar/ Asst. Registrar/ Superintendent, Examination Branch, UOM, Mysuru.
6. The PA to Vice-Chancellor/Registrar/Registrar (Evaluation), University of Mysore, Mysuru.
7. Office Copy.

Annexure-1

Cauvery College of Life Sciences and Management

Specialized PG programme by University of Mysore Credit Pattern(2022-2023)

M.Sc.(Reproductive Biology and Clinical Embryology (IVF)) (PG) Regulations and Syllabus

The credit pattern for Assisted Reproduction and Embryology is similar to the University's choice based credit system. However, the syllabus does not have soft core and open elective papers. All subjects are mandatory. The total credit for course is same as that of PG programme of the University.

Following shall be the minimum and maximum credits per semester.

The credit pattern is lecture (L); Tutorial (T); Practical (P); (L:T:P) Pattern.

Lecture: One hour session of theory class per week in a semester is 1 credit.

Tutorial and practical: Two hour session of tutorial or practical per week in a semester is 1 credit .

One semester period: 16 weeks of teaching and learning.

Duration of semester: 20 weeks that includes semester end examinations.

- A Candidate can enroll for a minimum of 19 credits per semester.
- A Candidate has to earn a minimum of 76 credits for successful completion of a master Degree.

Continuous assessment pattern:

Continuous assessment	Time duration	Marks		Minimum 30% and an aggregate of 40% to declare pass
		Max	Min	
C1	1 week to 8 weeks	15	4.5	
C2	9 weeks to 16 weeks	15	4.5	
C3	Complete 16 weeks	70	21	

Eligibility for Admission: Students of Bachelor of Science Degree (B Sc) from any UGC recognized Universities with life Science subjects or any equivalent bachelor degree/MBBS/BDS/BAMS/BHMS/B.Pharma/BSc Nursing/Engineering with sciences (Biotechnology and life science related) and Students from foreign nationals are also eligible, subjected to eligibility from University of Mysore. Admissions will be done as per University of Mysore norms.

SEMESTER I**(20) CREDITS**

SLNO	TITLE OF THE PAPER	CREDIT PATTERN			TOTAL CREDITS
		L	T	P	
1.1	Cellular biology & biophysical techniques	2	0	0	2
1.2	Human Reproductive biology	4	0	0	4
1.3	Andrology Laboratory Techniques-1	0	0	6	6
1.4	Andrology laboratory Techniques-2	0	0	8	8
		6	0	14	20

SEMESTER II**(20) CREDITS**

SLNO	TITLE OF THE PAPER	CREDIT PATTERN			TOTAL CREDITS
		L	T	P	
2.1	Genetics & Reproductive Endocrinology	2	0	0	2
2.2	Assisted Reproduction	2	0	0	2
2.3	In-Vitro Fertilization techniques	0	0	8	8
2.4	Clinical Embryology	0	0	8	8
		4	0	16	20

SEMISTER III**(20) CREDITS**

SLNO	TITLE OF THE PAPER	CREDIT PATTERN			TOTAL CREDITS
		L	T	P	
3.1	Infertility Counseling	2	0	0	2
3.2	Human Gametes and Embryo culture System	2	0	0	2
3.3	Micromanipulation in ART	0	0	8	8
3.4	Cryobiology and Techniques	0	0	8	8
		4	0	16	20

SEMISTER IV**(20) CREDITS**

SLNO	TITLE OF THE PAPER	CREDIT PATTERN			TOTAL CREDITS
		L	T	P	
4.1	Preimplantation Genetic Screening (PGS)	2	0	0	2
4.2	Quality control and risk management in ART	4	0	0	4
4.3	Writing and presentations skill	0	4	0	4
4.4	Project Work	0	0	10	10
		4	2	10	20

Semester1

(20Credits)

Paper1.1 CellularBiologyandBiophysicalTechniques

(2credits-2hours/Week)-32Hours

UnitI.CellBiology

Basic Characteristics of the Cell:Diversity of cell size and shape, Cell theory,Structure, organization and composition of prokaryotic and eukaryotic cell. Structureandfunctionsofcellorganelles,Celldivision-mitosisandmeiosis.Plasmamembrane

-structureandfunctions,**chromosomes**-TypesofChromosomesandmolecularanatomy of eukaryotic chromosomes – nucleosomes, organization and significance ofpolyteneandlampbrushchromosomes,Numericalchangesinchromosomes.Moleculare ventsofcelldivisionandcellcycle,regulationofcellcycle. **8Hours**

UnitII.Biochemistry

Structures of atoms, molecules and bonds, chemical foundations of biology, pH, pK,acids, bases and buffers, Covalent and non-covalent interactions, Vander – Wallforces, electrostatic and Hydrogen bonding and hydrophobic interactions, Laws ofthermodynamics,

Classification, structure and Properties of carbohydrates, amino acids and proteins,and lipids.Nucleic acids- Structure and properties- Bases, Nucleosides, Nucleotides,Polynucleotides. Structure of double stranded DNA (B, A, and Z-DNA. Types ofRNAsandtheirbiologicalsignificance. **8Hours**

UnitIII.BiophysicalTechniques

Introduction to Biophysics: Scope of Biophysics, Interaction of living and non-livingmatters,chemicalfoundationsofBiophysics.

Microscopic techniques: Principle and application of light, Phase contrast, Dark field,Fluorescencemicroscopy,ScanningandTransmissionElectronMicroscopy.

Chromatographytechniques:Principleandapplicationofpaperchromatography,TLC,Gelfiltration,chromatography,Ion–Exchangechromatography,Affinitychromatography,Gas-liquidchromatography,HPLC. **8Hours**

Centrifugation: Principle, techniques of preparative, analytical and ultracentrifugesElectrophoresisandBlottingTechniques,PrincipleandapplicationofPAGE, SDS-PAGE,Southern,NorthernandWestern blottingtechniques.

Spectroscopic techniques: Electromagnetic spectrum of light, Beer-Lambert's law,PrincipleandapplicationofUV,VisibleandIRspectroscopy,Fluorescencespectroscopy ,NMR, Atomic absorption spectroscopy, Massspectroscopy, Flowcytometry.

Radioisotopetechniques:Natureofradioactivity,detectionandmeasurement.Auto radiographyanditsapplications,BiologicalimportanceofLasers,MicrowavesandRadiations.

8hours

SEMESTER I

Paper 1.2 Human Reproductive Biology

(2 credits- 2 hours/Week)-32 Hours

UNIT 1

16 Hours

- Biomedical background of Bourn Hall Clinic
- The Bourn Hall story
- Gonadal changes from birth to puberty, Puberty and its timing, Environmental factors and puberty, Puberty and Psychological adjustment
- Anatomy of male and female reproductive systems
- Gamete biology: Spermatogenesis and oogenesis
- Andrology: Detailed Composition of seminal plasma,
- Semen Examination; Introduction, sample collection methods, sample collection for diagnostic or research purposes
- Sterile collection of semen for assisted reproduction and microbiological analysis
- Sample collection at home, collection of semen by condom
- Safe handling of specimens
- Initial examination: Liquefaction, semen viscosity, semen appearance, semen volume and PH
- Initial microscopic examination: Thorough mixing of the sample, making wet preparation,
- Cellular elements other than spermatozoa
- Sperm motility: categories of sperm movements, preparation and assessment of sperm motility
- Sperm vitality test: Using eosin-nigrosin, eosin alone and hypo-osmotic swelling test.
- Sperm Concentration estimation: Types of counting chambers
- Sperm Morphology assessment: Preparation of sperm smear, assessment of sperm morphology, staining procedures for sperm morphology
- Assessment of specific sperm defects
- Assessment of sperm leukocytes in semen
- Assessment of immature germ cells in semen

- Biomedical assay for accessory sex organ function: measurement of fructose and zinc in seminal plasma
- Ultrastructure of spermatozoa,

UNIT 2

16 Hours

- Male and female reproductive system: congenital or developmental disorders associated with primary testicular disorder,
- Klinefelter Syndrome, cryptorchism.
- Varicocele, infection, Drug and radiation
- Sperm transport disorders
- Poor semen parameters
- Causes of female infertility: Failure to ovulate, Problem in menstrual cycle, infection, failure to mature egg properly, endometriosis,
- Primary ovary insufficiency,
- Uterine anomaly and fibroid.
- Infertility trends worldwide, infertility in India
- Various Environmental factors effect on male and female infertility
- Clinical examination of male and female
- Female infertility: Disorders of female reproductive system, Fallopian tube block, hydrosalpinx
- Polycystic ovary syndrome (PCOS)
- Ovulation and anovulation
- Ovarian stimulation protocol

SEMESTER I

Practical paper-1.3

(6 Credits -

12 hours/week x 16 weeks) Paper title: Andrology laboratory techniques-1

1. Setting up of an IUI Laboratory,
2. Equipment and safety: Basic supplies needed in an Andrology laboratory,
3. Potential biohazards in Andrology laboratory, safety procedure,
4. Safety Procedure of laboratory Equipments, precautions when handling liquid nitrogen.
5. Brief Account of equipments: laminar airflow, various types of microscope, stereo zoom microscope, inverted microscope, Incubator used for IUI/dry bath, Centrifuge unit, Refrigerator, Makler chamber, Neuber chamber, sperm concentration
6. Use of CASA to sperm morphology assessment
7. Use of CASA to assess sperm motility,
8. Sperm preparation: Introduction, Choice of method, efficiency of sperm separation from seminal plasma and infectious organism, simple washing procedure, Direct swim-up, Discontinuous density gradient,
9. Preparation of HIV infected semen sample
10. Preparation of testicular and epididymal spermatozoa
11. Preparation of retrograde ejaculation sample
12. Antisperm antibody test
13. Biomedical waste Management

SEMESTER I

Practical paper-1.4

(8Credits -16hours

/weekx16weeks)Paper title:Andrology laboratory techniques-II

1. Semen examination with Kruger's criteria
2. Semen cryopreservation protocol: standard procedure, modified freezing protocols for poor semen samples, labelling of straws and record
3. Assessment of sperm chromatin: aniline blue test, acridine orange test, sperm chromatin structure assay (SCSA), Comet assay and TUNEL assay.
4. Quality control in the Andrology laboratory
5. Assessment of the Acrosome reaction
6. Assessment of nuclear decondensation of sperm and other functional test
7. Measurement of reactive oxygen species generated by leukocytes and sperm suspension
8. Sperm survival test
9. Advanced types of sperm preparation for ART
10. Mouse reproductive anatomy
11. Mouse reproductive physiology
12. Extraction of the oocytes from the ovary (Slaughterhouse)

Semester II

(20Credits)

Paper2.1:GeneticsandReproductive Endocrinology

(2Credits -2hours/week)-32Hours

UNIT1

16Hours

- Reproductive endocrinology: Hormones and their function, principles of hormone assay, various equipment to do hormone assay,
- Immunoassay, Hypothalamic pituitary function, thyroid hormones,
- Hirsutism primary and secondary amenorrhea.
- Male and female Endocrinology
- Hormonal regulation of spermatogenesis and ovarian cycle
- Luteal phase defect, Physiology of implantation.

UNIT2

16Hours

- Basic genetics of the cell,
- Chromosomal variations, Chromosomal syndromes, Genetic basis of evolution.
- Gene regulation: Translation, transcription, Expression, imprinting
- Basic Genetics: Genotype Phenotype, Mendelian inheritance pattern,
- Human karyotyping,
- DNA chromatin and Chromosomes,

- Gene Mutation and its effect,
- Epigenetics,
- Monogenic diseases, Chromosomal abnormalities numerical and structural.
- Human genome Project

SEMESTER II

Paper 2.2: Assisted reproduction

(2 Credits - 2 hours/week) - 32 Hours

UNIT 1

16 Hours

- History of assisted reproduction
- Gonadotropins in ART; Follicle stimulation hormone (FSH), Estradiol, progesterone and luteal hormone
- Removal of hydrosalpinges to improve IVF outcome,
- Ovarian reserve test, by AMH
- Antral follicle count

UNIT 2

16 Hours

- Ovarian endometriomas and IVF outcome.
- Uterine cavity abnormality and IVF outcome,
- Various kinds of sperm retrieval technique for ART
- Ultrasound in ovarian stimulation and follicle monitoring
- Ovarian hyperstimulation syndrome
- Oocyte markers of competence: Nuclear maturity
Cytoplasmic maturity
Polar bodies
Zona Perleucida
Cumulus cells

SEMESTER II

Paper 2.3: In-Vitro fertilisation techniques (IVF techniques)

Practical paper-1

(8 credits-16 hours/week x 16 weeks)

1. Preparation for IVF procedure: Going through patient file to know about kind of stimulation protocol, Gonadotrophins used.
2. previous history IVF if patient underwent IVF elsewhere
3. Culture Medium in ART: Media preparation for ART, detailed account of culture medium, sperm survival test, media preparation for intrauterine insemination (IUI) and IVF-ET
4. Method of fertilization, number of good quality oocytes, fertilization and number of embryos, and quality of embryos available.
5. Preparation for the IVF procedure: Medium aliquoting into sterile tube, Dish preparation for IVF and ICSI,
6. IVF witnessing
7. Dish preparation for IVF: close and open culture system
8. Sperm concentration calculation for IVF
9. Insemination of IVF droplet with sperm and co-incubation
10. Follicular fluid screening
11. Identification of oocytes, washing and pre-incubation
12. Assessment of Oocyte quality, IVF and fertilization check
13. Pronucleus grading
14. Fragmentation and grading the embryo

SEMESTER II

Paper 2.4: Clinical Embryology

Practical paper-2

(8 Credits -16 hours/week x 16 weeks)

1. Setting up of ART Unit with various facilities and detail Account on embryology laboratory and equipments.
2. Quality control, quality assurance and troubleshooting in IVF Lab
3. Equipment to control air quality in Embryology laboratory
4. Sterilization methods
5. Good laboratory practice (GLP)
6. Tissue culture techniques
7. Preparation of standard operation protocol for all procedures in the IVF Laboratory.
8. Culture system: Open and close culture advantages and disadvantages
9. Preparation for follicular fluid aspiration, culture medium disposables
10. Insemination of processed sample, Conventional IVF,
11. short term insemination and long term insemination, fertilization check, observation of fertilized Oocyte till Blastocyst development.
12. Cleavage stage embryo grading
13. Blastocyst grading
14. Blastocyst culture advantages and disadvantages

Semester III

(20Credits)

Paper3.1:Infertilitycounseling

(2Credits -2hours/week)-32Hours

UNIT1

16Hours

- Basicsofcounseling
- Infertilitycounseling
- Psychologyofinfertility
- Medicalaspectsoftheinfertilityforthecounselor
- Crossculturalissuesininfertilitycounseling
- Psychologicalevaluationoftheinfertilitycouple
- Evidencebasedapproachtointerfertilitycounseling
- Individualcounselingandpsychotherapy
- Counselingtheinfertilecouple

UNIT2

16Hours

- GroupapproachtointerfertilityCounseling
- Sexualcounselingandinfertility
- Geneticcounselingandtheinfertilepatient
- RecipientcounselingforDonorinsemination
- RecipientcounselingforEGGdonation
- Embryodonationcounseling
- Ethicalaspectsofinterfertilitycounseling
- LegalissuesininterfertilityCounseling
- AssistedreproductivetechnologyandtheimpactonChildren

Paper3.2:HumangameteandEmbryoculturesystem

(2Credits-2hours/week)-32Hours

UNIT1

16Hours

- Historicalbackgroundofgametesandembryoculture
- UtilityofanimalmodelsforHumanembryocultureMediacomposition
- Saltsandosmolarity,energysourceandmetabolism,

- amino acids and cellular homeostasis,
- Macromolecules and embryogrowth,
- antioxidant chelator and cellular function,
- pH and buffers, Growth factors.

UNIT 2

16 Hours

- Culture system: Single step and sequential
- Embryo co-culture
- Low-Oxygen culture
- Culture system: Embryo density
- Culture system: air quality
- Culture system: mineral oil overlay
- Human embryo culture media comparison
- Embryo culture and epigenetic

Semester III

Paper 3.3: Micromanipulation In ART

(8 credits - 16 hours/week x 16

weeks) Practical-1

1. History of micromanipulation,
2. various kinds of micromanipulation unit,
3. detail Account of all micromanipulation unit,
4. Micro tool preparation equipments,
5. Hands on practice of various micromanipulation unit,
6. Micro tool alignment,
7. ICSI dish preparation
8. Brief Account on PVP and hyaluronidase
9. Oocyte denudation
10. Oocyte assessment
11. Sperm immobilisation with various technique

12. IntraCytoplasmicsperminjection
13. Assistedhatching:Zonadrilling,Zonathinning,Chemicalandlaserassistedhatching
14. Intra-Cytoplasmicmorphologicallyselectedsperminjection(IMSI)
15. PhysiologicalselectionofspermandintraCytoplasmicsperminjection(PICSI)
16. Spindleview(Polo-scope)
17. Timelapsevideomonitoringofdevelopingembryo

Semester III

Paper3.4:CryobiologyandTechniques(8credits-16hours/weekx16weeks)Practical-2

1. Historyofgametecryopreservation
2. Psychologicalandpsychosocialissuessurroundingspermandegbanking
3. Legalandethicalaspectsofgametebanking
4. Methodofspermretrievalandbankingincancerpatient
5. DetailedAccountofcryoprotectantforslowfreezingandvitrificationmethod
6. Dishpreparationforfreezing/vitrification
7. Dishpreparationforthawing/warming
8. Advantageanddisadvantagebetweenslowfreezingandvitrificationmethod
9. Oocyte/spermvitrification
10. Cleavagestageembryovitrification
11. Blastocystcollapseandvitrification
12. Troubleshootinginvitrification
13. Ovariantissuecryopreservation:Harvestingovary
14. Preparationandprocessingofovariancortex
15. Vitrificationofovariancortex

16. Storage of vitrified ovarian cortex

17. Warming of ovarian cortex

18. Various equipment used for slow freezing

Semester IV (20 Credits)

Paper 4.1: Preimplantation Genetic Screening (PGS)

(2 Credits-2 hours/week)-32 hours

UNIT 1

16 Hours

- History of PGS: Animal studies and preclinical work, development of human embryo biopsy
- Genetic basis of inherited technologies
- Prenatal screening and diagnosis
- Preimplantation embryo development
- Preimplantation genetics
- Clinical aspects of Preimplantation genetics

UNIT 2

16 Hours

- Polar body biopsy
- Cleavage stage embryo biopsy
- Blastocyst biopsy
- Preimplantation genetic diagnosis for infertility (PGS)
- Preimplantation genetic diagnosis for sex-linked disease and sex selection for non-medical reasons
- Genetic counseling.

Paper4.2:QualitycontrolandriskManagementinART

(4Credits-2hours/week)- 64Hours

UNIT1

32 Hours

- QualityandqualitymanagementinARTlaboratory
- Regulation,licensingandaccreditation
- RiskandRiskmanagementinARTlaboratory
- Qualityandriskmanagementtool
- Riskeducation/riskminimization
- Howdowemanage risk,thebenefitofriskmanagement
- DevelopingRiskmanagementprogramme
- Howaredoingbenchmarking
- Humanresources

UNIT2

32Hours

- Parameterstorunsuccessfullaboratory
- MitochondrialreplacementtherapyinART
- ProteomicsandMetabolomics
- ICMRguidelineforARTunit,andregulationinassistedreproduction
- SurrogacyBill
- PCPNDAct
- Regulationandethicsinclinicalpractice
- Gameteandembryodonation
- RegulationofARTBank

Paper4.3:Writingandpresentationskill**(4credits)**

Students will select the appropriate topics related to ART under the guidance of teacher and prepare the powerpoint presentation. Prepared PPT will be presented during monthly seminar.

Paper4.4:PROJECTWORK**(10credits)**

Students pursuing M.Sc in Assisted Reproduction and Embryology Degree course is required to carry out work on a selected research project under the guidance of postgraduate teacher. This is to train a post graduate student in research methods and techniques. Project work includes identification of a problem, formulation of a hypothesis, search and review of literature, getting acquainted with recent advances, designing research study, collection of data, data analysis and comparison of results and finally drawing conclusions.

The projects should be written under the following headings

- Introduction
- Aims and objective of the study
- Review of literature
- Material and Methods
- Results
- Discussion
- Conclusion
- Summary
- Tables
- Annexure

Four copies of the project prepared shall be submitted to the department/university before the final examination date notified.

Scheme of examination

C3 Total marks

70 marks

Part-1 Simple answer questions (1 to 12 numbers)

2x10 Questions (12 questions)

20

marks Answer any ten of the following

Part-2 Descriptive questions. (13 to 20)

5x6 Questions

30 Marks

Answer any six of the following

Part-3

10x2 Questions

20 Marks

Write Long answer questions (21 to 24)

Answer any two of the following

RECOMMENDED READING BOOKS

Sl.No	NameOfTheBook	Authors	EditionAnd Year
1	AndrologyLaboratoryManual	AshokAgarwal,KaminiARao,MSSrinivas.	2010
2	APracticalGuideToSelectingGametesAndEmbryos	MarkusMontag.	2014
3	A Practical Guide To Basic Laboratory Andrology	ChristopherLRBarratt ,David Mortimer , JoseAntonio Castilla ,JuanG.Alvarez,LarsBjorn dahl.	2010
4	A WorkbookOnHumanSpermatozoaAndAssistedConception	AshokAgarwal,Sonia Malik.	2012
5	BiennialReviewOfInfertility	BartCFauser,CatherineRacowsky , Douglas TCarell,PeterNSchegel.	2013
6	Biochemistry	DavidRawn,J.	1989
7	Biochemistry	Voet,D.AndVoet,J.G.	1999
8	Biochemistry-The Chemical Reactions OfLivingCells-Vol-2.	DavidE.Metzle	1977
9	BiophysicalChemistryPart-2	CantorAndSchmmel.	1980
10	Biophysical Chemistry-Principles AndTechniques.	UpadyayaAndUpadyaya.	2003
11	Chemistry–AnIntroductionToGeneral,OrganicAndBiologicalChemistry.	KarenC.Timberlake	1999
12	EmbryoCulture:MethodsAndProtocols,	GaryDSmith,JasonE.Swain,ThomasB.Pool.	2013
13	EmbryoTransfer	GautamNAllahbadia	2008

14	Handbook Of Human Oocyte Cryopreservation	Eleonora Porcu, Patrizia Maria Ciotti, Stefano Venturoli	2013
15	Harrison's Endocrinology	J. Larry Jameson	2010
16	Harper's Review of Biochemistry, (1997)	Murray et al.	1997
17	Human Assisted Reproductive Technology: Future Trends in Laboratory and Clinical Practice.	Botros R. M. B. Rizk, David K Gardner, Tommaso Falcone.	2011
18	Infertility Counseling - A Comprehensive Handbook For Clinicians.	Linda Hammer Burns, Sharon N Covington	2006
19	Lehninger: Principles of Biochemistry	David L Nelson, Michael Cox	2015
20	Male Infertility, 2 nd Edition,	T. B. Hargreave.	1997
21	Medical Laboratory Technology Part-	SK. Mizanur Rahman.	2009
22	Molecular Biology of the Cell.	Bruce Alberts	2002
23	Netter's Atlas of Human Embryology	Larry R. Cochard	2002
24	Quality and Risk Management in the IVF Laboratory	David Mortimer and T. Mortimer	2008
25	Oocyte Biology in Fertility Preservation	S. Samuel Kim	2013
26	Sperm Banking - Theory and Practice	Allan A Pacey, Mathew J Tomlinson	2009
27	Textbook of IUI and ART	Hrishikesh Pai, Kinjal R Shah, Nanditha Palshetkar, Rishma Dhillon Pai.	2016
28	Textbook of Biochemistry with Clinical Correlations.	Thomas Devlin	1999
29	Textbook of In Vitro Fertilization and Assisted Reproduction: The Bourn Hall Guide to Clinical and Laboratory Practice. 3 rd Edition	Peter R Brinsden	2005
30	The Subfertility Handbook - A Clinicians	Gab Kovacs .	2011

	Guide, 2 ND Edition,		
31	Textbook Of Assisted Reproductive Technologies: Laboratory And Clinical Perspectives. 3 rd Edition	Ariel Weissman, Colin M Howles, David K Gardner, Zeev Shoham.	2009
32	Practical Manual Of In Vitro Fertilization.	Alex C. Varghese, Ashok Agarwal, Zolt Peter Nagy	2013
33	Practical Biochemistry; Principles And Techniques;	K. Wilson And J. Walker (1995) 4 th Edn.	1995
34	Preimplantation Genetic Diagnosis 2 nd Edition	Joyce Harper	2009